

1 **ISOLATION AND IDENTIFICATION OF DIESEL-DEGRADING BACTERIA FROM**
2 **OIL CONTAMINATED SOIL IN MANSEHRA, PAKISTAN**

3 **Zia-Ur-Rehman¹, Khakemin Khan², Shah Faisal^{1, 4}, Rasool Kamal¹, Shazad Ahmad¹,**
4 **Irfan¹, Mian Khizar Hayat⁴, Safia⁵, Inayat Ullah¹, Tanweer Kumar³**

5 ¹ Department of Microbiology, Faculty of Health Sciences, Hazara University Mansehra KPK
6 Pakistan.

7 ² Department of Chemistry Comsats Institute of Information Technology Abbottabad Pakistan
8 22060

9 ³College of Pastoral Agriculture Science and Technology, Lanzhou University, Lanzhou 730000,
10 P.R. China

11 ⁴MOE Key Laboratory of Cell Activities and Stress Adaptations, School of Life Sciences,
12 Lanzhou University, Lanzhou, Gansu 730000, China

13
14 ⁵ Department of Ophthalmology College of allied and vision science king Edward medical
15 university

16 * **Corresponding Author**

17 **Khakemin Khan (Corresponding Author)**

18 E-Mail khakemin_chemist@yahoo.com

19 Tel: +92-(313)-9076940

20

21 **ABSTRACT:** - The present work was conducted to isolate and identify bacteria from oil-
22 contaminated soil to evaluate their role in biodegradation of commercial diesel under laboratory
23 conditions. Diesel fuels are used by different vehicles, diesel generators and especially heavy
24 transport vehicles. Its manufacturing, transportation, utilization and disposal have the threat to
25 pollute the surrounding environment. Biodegradation is one of the biological processes to
26 remediate the pollutants. This is the cheaper and easy method as compared to other methods like
27 direct burning, land foaming and bioventing because diesel smoke has a carcinogenic effect.

28 In the current exploration ordinary bacterial species that degrade diesel were isolated from
29 selected areas of Mansehra. Six sites were selected and a total of 60 samples were collected.
30 These sites were coded according to their names. For the isolation of diesel degrading bacteria
31 soil samples were analyzed. The isolation of bacteria was done on nutrient agar, nutrient broth
32 and mineral salt medium (MSM), and IR spectroscopy was used to observe the degradation of
33 diesel. Twenty samples were found having bacterial growth. Further analysis showed that four
34 samples BHG, SHNK, TAK and KMOR had diesel degrading capability as these showed
35 positive results. The isolated bacteria were identified by morphological and biochemical features,
36 while the degradation of diesel was determined qualitatively by interpreting the intensity of the
37 peaks of IR spectroscopy. The intensity of the peaks in the spectrum of control was compared
38 with the intensity of the peaks in the spectrum of the test samples which indicated degradation of
39 diesel. Three bacterial species, namely, *Bacillus cereus*, *Bacillus subtilis* and *Pseudomonas*
40 *aeruginosa* were identified using Berge's manual method. The bacterial species were tested for
41 their capability to degrade commercial diesel presented at different concentrations i.e. 1000 ppm
42 and 5000 ppm.

43 The highest degradation capability of commercial diesel was exhibited by *Bacillus cereus*
44 followed by *Bacillus subtilis* and *Pseudomonas aeruginosa*.

45 The results indicate that *Bacillus cereus*, *Bacillus subtilis* and *Pseudomonas aeruginosa* have the
46 potential of *in situ* bioremediation of diesel-contaminated soils.

47 **Key words:** *Diesel, Biodegradation, Spectroscopy, Bacillus cereus, Bioremediation, Bacteria*
48 *pseudomonas auerginosa*.

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50 INTRODUCTION

51 Mansehra District is located at the eastern border of the Khyber Pakhtunkhwa Province, three
52 hours away from Islamabad and four hours from Peshawar.

53 In the current exploration diesel-degrading bacteria were isolated and identified. Diesel fuel, in
54 general, is any liquid fuel used in diesel engines. The most common is a specific fractional
55 distillate of petroleum fuel oil, but alternatives that are not derived from petroleum, such as
56 biodiesel, biomass to liquid (BTL) or gas to liquid (GTL) diesel, are increasingly being
57 developed and adopted. To distinguish these types, petroleum-derived diesel is called petro
58 diesel. Petroleum-derived diesel is composed of about 75% saturated hydrocarbons (primarily
59 paraffins including *n*, *iso*, and cycloparaffins), and 25% aromatic hydrocarbons (including
60 naphthalene and alkyl benzenes). The average chemical formula for common diesel fuel is
61 $C_{12}H_{23}$, ranging approximately from $C_{10}H_{20}$ to $C_{15}H_{28}$.

62 The commercial diesel oil is the mixture of normal, branched, cyclic and aromatic substances
63 having the characteristics of low water solubility, high adsorption and strong stability of aromatic
64 rings (Dean *et al.*, 2002). With the increase in world population the need of petroleum and diesel
65 fuel is increasing. Use of huge quantities and consumption of these products directly affects the
66 surrounding environment (Raven *et al.*, 1993). The common major global environmental
67 pollution is due to the combustion of diesel fuel, crude oil and petroleum compound. These
68 hydrocarbons are mostly used as primary source of energy. There is a need of large quantities
69 of fuel to run auto mobile and power industry and also used for home heating. During storage,
70 transportation and transformation leakage accidents the environment gets polluted (Watanabe,
71 2001; Surridge, 2007).

72 Huge hydrocarbon contaminants are caused by oil spills from marine accidents during shipping
73 and transport of crude oil (Ghosh *et al.*, 2006). Diesel fuel contaminants having very high
74 carcinogenicity and mutagenicity serve as strong toxicants causing various human and animal
75 health problems (Boonehan *et al.*., 2000 ; Samante *et al.*., 2002). There are various methods for
76 cleaning ground environment from petroleum and diesel hydrocarbon. Among these methods
77 bioremediation is more effective and approachable method to remove hazardous substances,
78 than other commercially available techniques (Alexander, 1994;Ojo, 2006).The process of
79 bioremediation is nowadays used in many countries of the world to clean up our
80 environment from chemical contaminants (Skladany and Metting, 1992). Bioremediation is
81 an emerging field for investigators to present the biological treatment of environmental pollutant,
82 by means of microbes. There are numerous organisms which have the ability to utilize diesel
83 hydrocarbon as rich source of carbon and convert these organic contaminants to harmless
84 compounds (Atlas and Philp, 2005; Koren *et al.*, 2003).

85 Petroleum and diesel hydrocarbon degradation is not a new concept. It is 50 years old
86 phenomenon in which different bacteria having the ability to degrade petroleum and diesel
87 hydrocarbons have been isolated from contaminated soil, storage tanks and from oil spills
88 environment (Huy *et al.*, 1999).There are many microorganisms which have the ability to
89 degrade these pollutants from the environment and many scientists are working to get solution
90 for removing such kind of pollutants

91 Degradation of hydrocarbon in contaminated soil is a safe, competent, domestic and economical
92 method to degrade harmful contaminants (Mehrashi *et al.*, 2003). Natural population of microbes
93 promotes breakdown of petroleum and diesel hydrocarbon by converting hazardous compounds

94 into less toxic substances and remediation of contaminants from the environment is completely
95 approachable (Atlas *et al.*, 1981).

96 There are different types of bacteria that are capable of degrading diesel and petroleum
97 hydrocarbons. Only one bacterial species is unable to degrade hydrocarbon molecules. Among
98 naturally occurring bacteria *Pseudomonas spp* are able to utilize carbon as energy source because
99 they contain plasmids, and the relevant genes for degradation (Jawetz *et al.*, 1991). Different
100 bacterial species are isolated successfully from Soil samples from oil contaminated sites.
101 Bacteria of the genus *Staphylococcus* were found as the major degrading microorganisms
102 (DeRito *et al.*, 2005). It can be postulated that gram positive organisms are in the family of
103 such microorganisms that take a part in breakdown of complex hydrocarbon. Some Gram-
104 positive bacteria, mostly *Rhodococci* have the ability to break alkanes (Kaplan and Kitts,
105 2004).

106 The main objective of this paper is to clearly describe the bacteria isolated from the oily
107 contaminated soil and also to identify those bacterial species which may have potential to be
108 exploited as agents for degradation of commercial diesel.

109 **MATERIALS AND METHODS**

110 **Soil sample collection**

111 Soil samples were collected from oil contaminated soils around the storage tanks inside the
112 filling stations, bus/trucks stand, car wash station and auto workshops from different localities in
113 District Mansehra, such as General bus stand, Shinkiari, Takkra, etc.

114 The soil was contaminated with diesel and other petroleum fuel spilled and at least 30 soil
115 samples approximately 20 g were collected randomly from the top soil layer up to 10-30 cm in
116 depth using a hand auger followed by bulking in the selected areas. After removal of surface

117 litter, stone and big soil debris, the samples were passed through a 2 mm sieve and stored at 4 °C
118 in sterile plastic bags. Samples were immediately transported to the laboratory for further
119 analysis.

120 **Enrichment medium**

121 All microbial enrichment was performed in the mineral salt medium (MSM). This medium was
122 used for the growth of such bacteria having the capability to degrade petroleum hydrocarbon.
123 The enrichment medium is also called Basal Salt medium (BSM) and their composition are
124 mentioned below.

125 (g/L): NaNO₃ (7); K₂HPO₄ (1); KH₂PO₄ (0.5); KCl (0.1); MgSO₄.7H₂O (0.5); CaCl₂ (0.01);
126 FeSO₄.7H₂O (0.01); H₃BO₄, (0.25); CuSO₄.5H₂O (0.5); MnSO₄. H₂O (0.5) and ZnSO₄.
127 7H₂O (0.7), containing 25 to 100 mg of yeast extract or tryptone (Mercade *et al.*, 1996).

128 All the ingredients of the medium were dissolved in distilled water and shake properly.

129 The enrichment medium was prepared in 1000 ml flask. The medium was then autoclaved at
130 121°C for 15 minutes for sterilization and the pH of the medium was adjusted to 7.3 with a drop
131 of HCl or NaOH.

132 **Cultivation of diesel fuel-degraders**

133 The mineral salt medium (MSM) was used for cultivation of diesel degrading bacteria as
134 describe by Mercade *et al.*, (1996).

135 Bacterial enrichment was carried out in 250 mL Erlenmeyer flasks containing 100 mL of MSM.

136 **First transfer**

137 From the prepared Enrichment medium 100 ml media was transferred to 250 ml Erlenmeyer flask
138 with commercial diesel as sole source of carbon and energy. One gram of the diesel

139 contaminated sieve soil was added to 100 ml of the enrichment medium and incubated at 35 °C
140 by using orbital incubator (120 rpm) for the period of 15 days.

141 **Second transfer**

142 After the completion of initial cultivation one ml of enriched culture was transferred into newly
143 prepared enrichment medium and incubated at the same temperature in the orbital shaker (120
144 rpm) for the 15 days. The third transfer was same as second as describe by Singh and Lin,
145 (2008).

146 **Isolation of Suspected Diesel Degraders**

147 After the completion of enrichment process 1 ml culture was serially diluted to six levels (10-
148 fold) were made in sterile water from the third enrichment culture. For isolation of suspected
149 diesel degraders 100 µl of diluents was streak on nutrient agar plates from the last dilution (6th
150 level) and the plates were covered with 100 µl of diesel oil before streaking and incubate at 30°C
151 for 48 hours. The nutrient agar (NA) and broth was used for isolation, numeration and
152 maintenance of pure cultures of diesel degraders.

153 **Bio-degradation Analysis**

154 Bio-degradation of commercial diesel fuel was performed in different low 1000 ppm and high
155 5000 ppm concentration as the sole source of carbon and energy.

156 A series of twenty two Erlenmeyer flasks of 300 ml capacity were used for this experiment. Out
157 of these twenty two flasks ten flasks were used for 1000 ppm and ten flasks were used for 5000
158 ppm and two flasks each was used for reference/controls. Each flask contained 100 ml of 8 g/l of
159 nutrient broth and was sterilized by autoclaving (121 °C for 15 min). After autoclaving 100 ml of
160 1000 ppm diesel solution from stock solution was added in to each of the 11 flasks and similarly
161 100 ml from 5000 ppm diesel solution was added to the next series of 11 flasks.

162 The suspected diesel degrading bacteria in nutrient broth were shaken and the flasks were
163 inoculated by transferring 2 ml of pre-culture (about 3×10^8 cells/ml) of bacteria and initial pH
164 was adjusted to 7.3. The experimental flasks were incubated under aerobic condition in an orbital
165 incubator (120 rpm) at temperature (30 °C) for 20 days.

166 **Qualitative Determination of degraded diesel**

167 Samples were randomly removed for qualitative determination by Fourier transform infrared
168 spectroscopy (FTIR) Version 1.30 Shimadzu Corporation and bacterial augmentation were
169 monitored by viable counts on nutrient agar plates (Amund and Igiri, 1990).

170 **Identification of Isolated Diesel Degradars**

171 After the completion of degradation analysis the suspected diesel degrader's growth was
172 monitored by viable counts on nutrient agar plates and then the diesel degraders were identified
173 by various methods.

- 174 i. Morphological identification
- 175 ii. Biochemical identification

176 **Morphological identification**

177 Basic morphological identification was performed including colony morphology, shape and
178 color of colony and Gram-reaction to identify the bacterial diesel degraders (Balows *et al.*,
179 1992). For colony morphology, shape and colour of samples were examined through compound
180 microscope and the results were recorded.

181 **Biochemical identification**

182 Biochemical identification is based on biochemical tests including catalase, coagulase, urease,
183 oxidase activities, Simmons citrate, methyl red voges prosker (MRVP) and indol productions
184 were checked as recommended by Smibert and Krieg (1994). Triple Sugar Iron Agar is used for

185 the differentiation of microorganisms on the basis of dextrose, lactose and sucrose fermentation
186 and hydrogen sulfide production (Kligler, 1918).

187 **RESULTS AND DISCUSSION**

188 *Properties of soil*

189 The physico-chemical properties of the soil samples like texture and pH along with their codes
190 are presented in Table 1.

191 *Cultivation of diesel fuel-degraders*

192 The cultivation of diesel fuel degraders were carried out in 250 mL Erlenmeyer flasks containing
193 100 mL of MSM and few drops of commercial diesel as sole source of carbon and energy as
194 describe by Mercade *et al.*, (1996). One gram of the diesel contaminated sieve soil was added to
195 100 ml of the enrichment medium and incubated at 35 °C by using orbital incubator (120 rpm)
196 for the period of 15 days. After the completion of initial cultivation 5 ml of enriched culture was
197 transferred into newly prepared enrichment medium and incubated at the same temperature in
198 orbital shaker (120 rpm) for the 15 days. After the completion of 2nd transfer of bacterial
199 cultivation finally transfer 2 ml of enrichment culture in to the third newly prepared medium and
200 incubated at 35 °C by using orbital incubator (120 rpm) for the period of 15 days, Singh and Lin,
201 (2008).

202 *Isolation of Diesel Degradars*

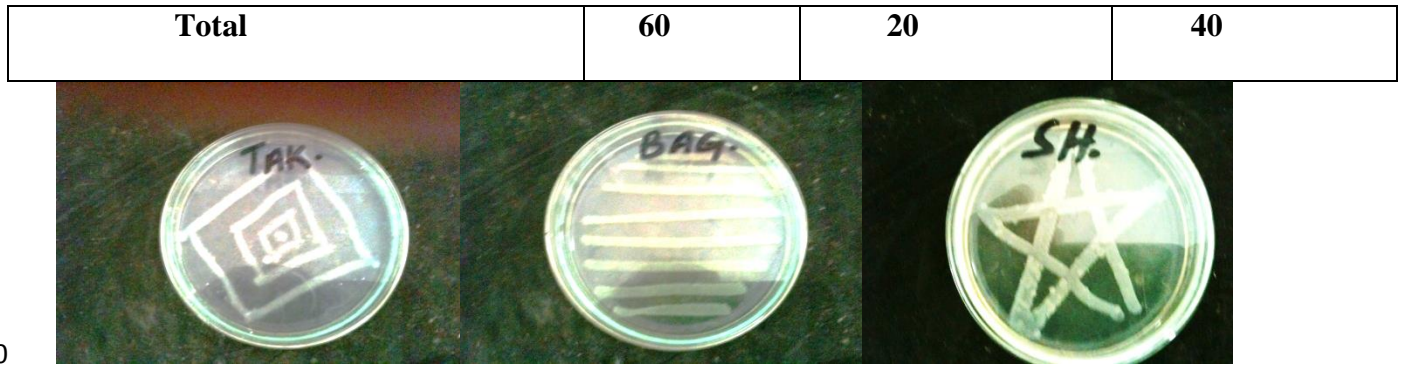
203 After the completion of cultivation process, a 10-fold serial dilution of the culture up to 6th level
204 was made and 100 µl of diluents was streak on nutrient agar plates from last dilution as describe
205 in materials and methods.

206 A total of 30 samples were run, 18 samples shows growth. Five samples from each selected area
 207 were collected. From Shinkiari and Bhagwar areas 10 samples were collected and run and only 6
 208 samples shows growth. From Athar shesha 5 samples were collected and only 2 samples shows
 209 growth and then the total of 5 samples were collected from Takkra and only 4 samples shows
 210 growth. From Kalgan moare and Mansehra General Bus stand a total of 10 samples were
 211 collected and only 2 and 4 samples show growth respectively. The details of the total samples
 212 and the number yielding bacterial growth are given in Table 2.

213 The samples which showed 40 to 112 colonies were examined per plate on the basis of colony
 214 growth appearance were inoculated in nutrient broth in order to sub cultured for obtaining pure
 215 cultures that were further use for degradation and identification process. These colonies have
 216 distinct features such as rods and become wrinkled, convex or flat surface and ranging from
 217 brownish, greenish and greenish, yellowish were selected. All samples demonstrated a successful
 218 growth in nutrient broth as viewed by turbidity of the cultures, as shown in figure 1.

219 Table No 2: Distribution of samples yielding or not yielding bacterial growth by sites.

S.No	Origin of samples	Samples code	Total no of samples	Samples yielding growth	Samples not yielding growth
1	Bhagwar	BHG	10	3	7
2	Shinkiari	SHNK	10	6	9
3	AhtarShesha	ASM	10	2	8
4	Takkara	TAK	10	4	6
5	Kalgan Moare	KMOR	10	2	8
6	Mansehra Gen bus stand	MGBS	10	3	7



220

221 Figure 1. Petri plates showing growth of bacterial strains after 5 days incubation.



222

223 (a)

(b).

(c)

224 Figure 2. Significant growths in nutrient broth of collected soil samples were shown.

225 **Degradation analysis of commercial diesel by isolated bacterial species**

226 Degradation of commercial diesel by isolated bacterial species was confirmed by Fourier
227 transform infra-red (FTIR). Commercial diesel was presented to the test organisms in two
228 concentrations 1000 ppm and 5000 ppm for 15 days as described in materials and methods.

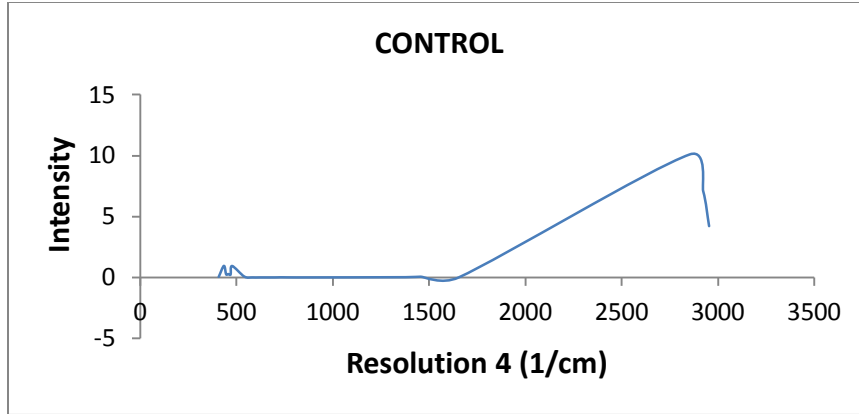
229 The results of FTIR analysis showed that the introduction of bacterial species enhanced the
230 degradation of commercial diesel as compared to the samples that were not supplemented with any
231 bacterial species. Commercial diesel degradation was analyzed by using FTIR. First of all a
232 control sample was run having commercial diesel concentration of 1000 ppm and 5000 ppm in the
233 absence of bacterial species. It showed IR spectroscopic peak at 2852.72 cm^{-1} , 2922.16 cm^{-1} ,
234 2853.02 cm^{-1} corresponds to the stretching vibration of hydrogen and carbon bonds which was

235 recorded on a graph paper (Figure 2). The intensity of the above peaks in the spectrum of control
236 was compared with the intensity of the peaks in the spectrum of the test samples. This response
237 was used as standard/reference for comparison with results of test samples offered diesel at a
238 concentration of 1000 ppm.

239 To validate the degradation of diesel in 10 samples with 1000 ppm concentration were tested in
240 FTIR separately. The 1000 ppm concentration control/standard intensity of peaks at 2852.72 cm^{-1} ,
241 2922.16 cm^{-1} , 2853.02 cm^{-1} was 10.1, 7.11, 4.22 upon calculations the overall intensity was 21.42
242 which was high as compared with the intensity of a spectrum of the samples.

243 It has been found that only 4 samples showed degradation of commercial diesel as the IR
244 spectroscopic peaks at 2852.72 cm^{-1} , 2922.16 cm^{-1} , 2853.02 cm^{-1} have low intensity as compared
245 with standard/control. The sample taken from BHG showed degradation of diesel which has low
246 intensity which was 16.53 that indicates the degradation of diesel as it was different from 21.42 of
247 standard peak of diesel as shown in Figure 3 and table 3.

248 The samples taken from TAK and SHNK also showed degradation of diesel. The intensities of
249 these samples were recorded as 13.1, 18.89 and 17.05 respectively. The results of FTIR spectrum
250 peaks obtained on graph paper were shown in Figures 4, 5, and 6. The values of IR spectroscopic
251 peaks and the low intensity recorded in these samples were the clear signal of diesel degradation
252 compared with standard/control as shown in tables 4,5 and 6.



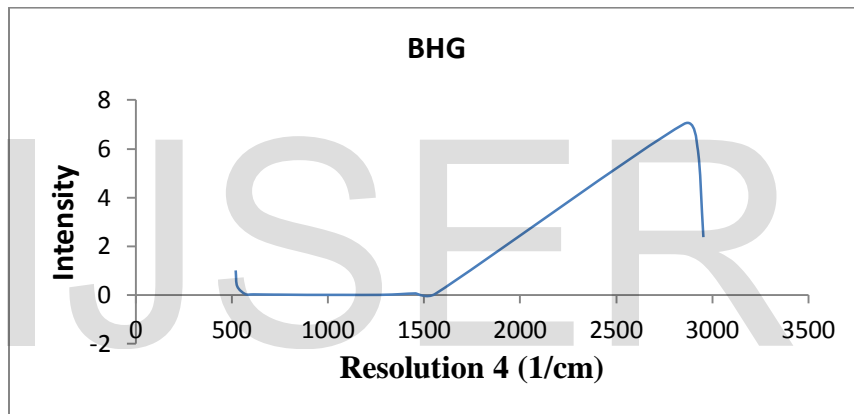
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254

Figure 2. Standard IR spectroscopic peak of Diesel without any bacterial species at 1000

255

ppm



256

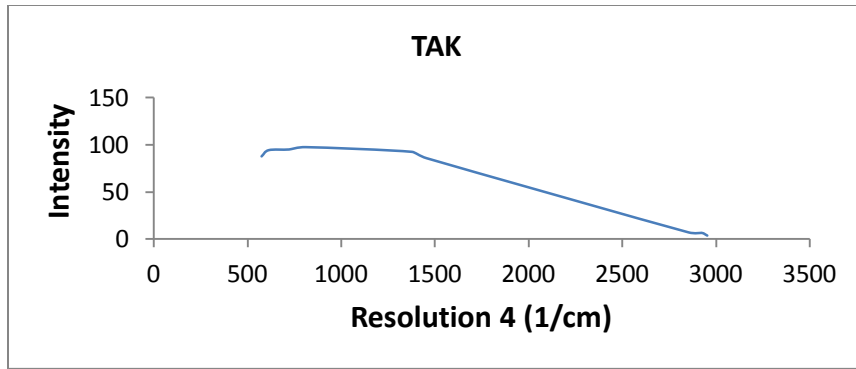
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Figure.3. IR spectroscopic peak of Sample BHG with *Bacillus cereus* at 1000 ppm

Intensity of control (1000 ppm)			Intensity of sample BHG (1000 ppm)	
S. no	Peaks	Intensity	Peaks	Intensity
1	2852.72	10.01	2852.72	7.02
2	2922.16	7.10	2922.16	6.08
3	2953.02	4.22	2953.02	2.43
Total		21.42	Total	16.53

258

Table 3. Intensity pattern of Sample BHG with *Bacillus cereus* at 1000 ppm



259

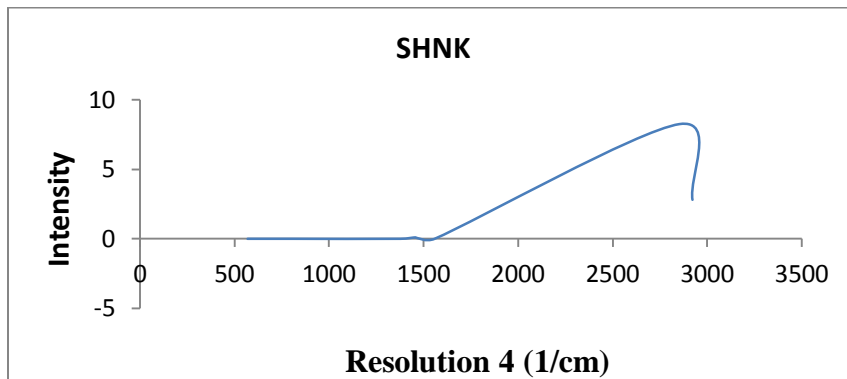
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Figure 4. IR spectroscopic peak of Sample TAK with *Bacillus subtilis* at 1000 ppm

Intensity of control (1000 ppm)			Intensity of sample TAK (1000 ppm)	
S. no	Peaks	Intensity	Peaks	Intensity
1	2852.72	10.01	2852.72	7.04
2	2922.16	7.10	2922.16	6.53
3	2953.02	4.22	2953.02	3.63
Total		21.42	Total	17.2

261

Table 4. Intensity pattern of Sample TAK with *Bacillus subtilis* at 1000 ppm



262

263

Figure 5. IR spectroscopic peak of Sample SHNK with *Pseudomonas aeruginosa* at 1000

264

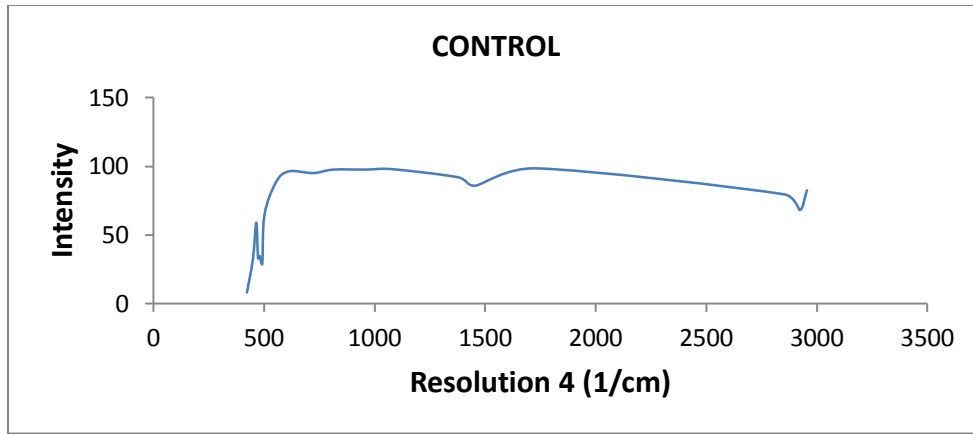
ppm

Intensity of control (1000 ppm)			Intensity of sample SHNK (1000 ppm)	
S. no	Peaks	Intensity	Peaks	Intensity
1	2852.72	10.01	2852.72	8.26
2	2922.16	7.10	2922.16	7.82
3	2953.02	4.22	2953.02	2.81
Total		21.42	Total	18.89

265 Table 5. Intensity pattern of Sample SHNK with *Pseudomonas aeruginosa* at 1000 ppm

266 In the second assessment the test samples were presented diesel in concentration of 5000 ppm. A
 267 control sample without any bacterial specie was evaluated in FTIR spectroscopy. Its peak was
 268 same i.e. 2852.72 cm^{-1} , 2922.16 cm^{-1} , 2853.02 cm^{-1} but different in the intensity 229.21 was recorded
 269 on a graph paper as shown in Figure 6.

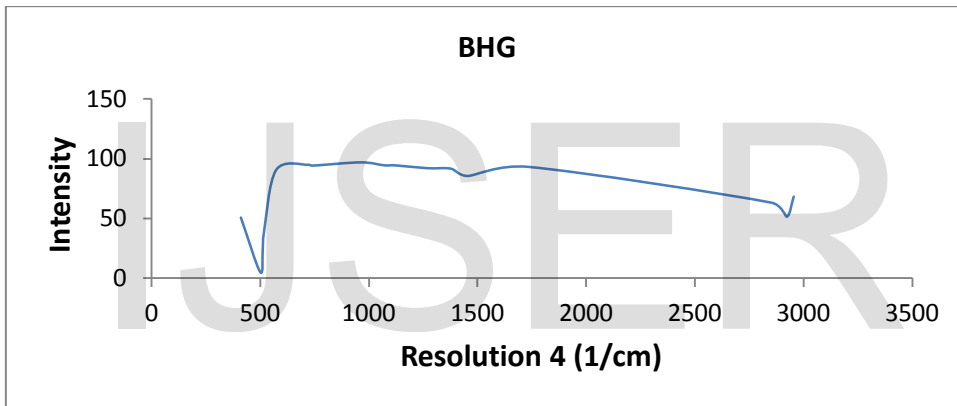
270 A comparative study of diesel degradation was carried out in 10 test samples incubated with
 271 diesel at 5000 ppm concentration. In these 10 samples only 3 samples, taken from BHG, SHNK
 272 and TAK showed degradation. The diesel degradation of these samples was tested in FTIR
 273 spectroscopy. The intensities of the peaks of these samples were recorded on a graph paper and it
 274 was found that the sample taken from BHG was low intensity i.e.182.94 while samples taken
 275 from SHNK and TAK have intensities of 162.51 and 187.68 respectively. The peaks of these 3
 276 samples evidently indicated the diesel degradation because the intensity of the control was high as
 277 compared with these samples as shown in Figures 7, 8 and 9 compared to the standard/control as
 278 shown in table. 6, 7 and 8.



279

280 Figure 6. Standard IR spectroscopic peak of Diesel without any bacterial species at 5000

281 ppm.



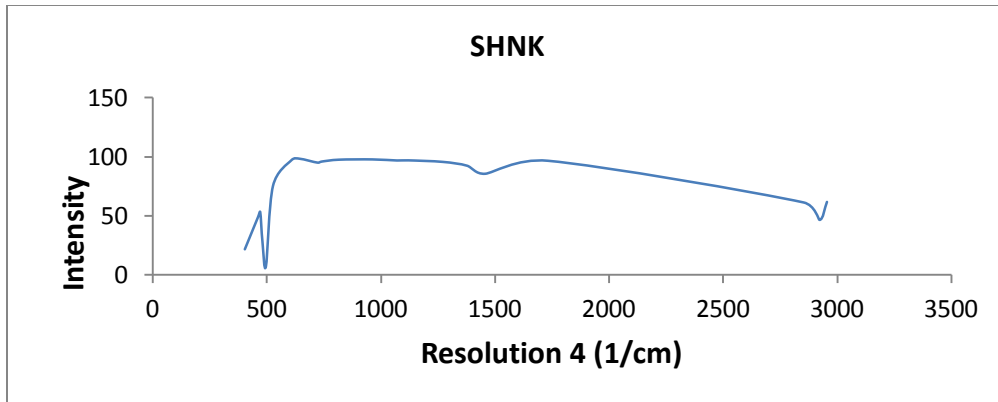
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283 Figure 7. IR spectroscopic peak of Sample BHG with at *Bacillus cereus* 5000 ppm

Intensity of control (5000 ppm)			Intensity of sample BHG (5000 ppm)	
S. no	Peaks	Intensity	Peaks	Intensity
1	2852.72	78.98	2852.72	63.22
2	2922.16	67.69	2922.16	51.43
3	2953.02	82.54	2953.02	68.29
Total		229.21	Total	182.94

284

Table 7. Intensity pattern of Sample BHG with *Bacillus cereus* at 5000 ppm



285

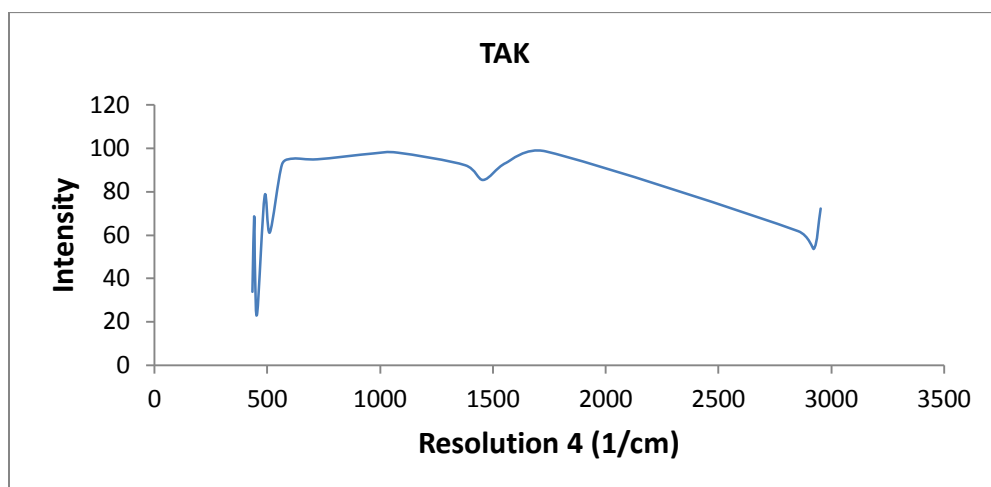
286 Figure 8. IR spectroscopic peak of Sample SHNK with *Pseudomonas aeruginosa* at 5000

287 ppm

Intensity of control (5000 ppm)			Intensity of sample SHNK (5000 ppm)	
S. no	Peaks	Intensity	Peaks	Intensity
1	2852.72	78.98	2852.72	61.22
2	2922.16	67.69	2922.16	46.43
3	2953.02	82.54	2953.02	61.86
Total		229.21	Total	169.51

288

Table 8. Intensity pattern of Sample SHNK with *Pseudomonas aeruginosa* at 5000 ppm



289

290 Figure 9. IR spectroscopic peak of Sample TAK with *Bacillus subtilis* at 5000 ppm

Intensity of control (5000 ppm)			Intensity of sample TAK (5000 ppm)	
S. no	Peaks	Intensity	Peaks	Intensity
1	2852.72	78.98	2852.72	61.83
2	2922.16	67.69	2922.16	53.61
3	2953.02	82.54	2953.02	72.24
Total		229.21	Total	187.68

291 **Table 9.** Intensity pattern of Sample TAK with *Bacillus subtilis* at 5000 ppm

292 **Identification of Isolated Diesel Degraders**

293 Four bacterial isolates showed positive results during diesel degradation analysis of the selected
 294 samples. These bacterial species were identified based on both morphological and biochemical
 295 identification techniques.

296 Different morphological characteristics were considered during identification. On the basis of
 297 these morphological features three bacterial species namely *Pseudomonas aeruginosa*, *Bacillus*
 298 *subtilis* and *Bacillus cereus* were tentatively identified. The morphological features of these
 299 strains are given in Table 10.

300 Table 10: Morphological features of bacterial strains

S. no	Sample code	<u>Growth samples characteristics</u>			Bacterial species identified tentatively
		Cell shape	colony colour	Gram test	
1	BHG	Smooth, circular	white-cream	Gram +ive	<i>Bacillus cereus</i>

2	SHNK	Rod-shaped	blue/green	Gram -ive	<i>Pseudomonas aeruginosa</i>
3	TAK	Rod-shaped	Brownish	Gram +ive	<i>Bacillus subtilis</i>

301
 302 The identification of the isolated bacterial species was further confirmed by using various
 303 biochemical tests. The results of these biochemical tests are given in Table 11.

304 Table 11: Identification and characterization of diesel-degrading bacteria using various
 305 biochemical tests

S. No.	Biochemical tests	<i>Bacillus cereus</i> (BHG)	<i>Pseudomonas aeruginosa</i> (SHNK)	<i>Bacillus subtilis</i> (TAK)
1	Catalase	+	+	+
2	Fluorescent Pigment	-	+	-
3	Oxidase	+	+	-
4	Indol	-	-	-
5	Methyl red	-	-	-
6	Motility	+	+	+
7	Simmons citrate	+	+	+
8	Urease	+	+	+
9	voges-proskauer	+	-	+

306

307 Table 12: Identification and characterization of diesel-degrading bacteria using TSI

308

Organism	Triple sugar iron			
	Slop	Butt	H ₂ S	GAS
<i>Bacillus subtilis</i>	R	Y	-	-3
<i>pseudomonas aureginosa</i>	R	Y	+	-
<i>Bacillus cereus</i>	R	Y	-	-3

309

310 Key: H₂S=hydrogen sulphide gas (blackening), R= red pink (alkaline reaction), Y=Yellow (acid
 311 reaction)

312 The results stated that Identification on the basis of biochemical test and TSI were confirmed that
 313 species characters were similar to *Bacillus subtilis*, *Bacillus cereus* and *pseudomonas auerginosa*
 314 as describe by Smibert and Krieg (1994).

315 **Discussion**

316 The present exploration was carried out to assess the status of biodegrading bacteria in soil
 317 samples collected from various localities in district Mansehra. The results of the current
 318 investigation evidently indicated that limited samples gave positive results. The ability of
 319 isolated bacterial species to degrade commercial diesel was also evaluated using IR
 320 spectroscopy. The method of biodegradation of important chemicals by microorganisms has
 321 gained much significance in the field of Biotechnology. Now several researches are being
 322 conducted in various parts of the world to isolate valuable biodegrading bacteria and fungi from
 323 soil or other sources and then use these species in environmental Microbiology.

324 Soil contaminated with petroleum hydrocarbon is the most probable resource to isolate diesel
325 degrading microbes. The degradation potential of isolated bacteria was examined. Disclosure to
326 rising point of diesel concentration was used to find out the resistance of isolated strain. The use
327 of microbes to degrade such health perilous compounds is the need of time. Several researches
328 are being conducted to build up techniques to isolate such kind of degrading bacteria from soil or
329 water and then use these strains on commercial basis to hygienic our environment from such
330 carcinogenic compounds.

331 Diesel is one of the most ordinary pollutants present in the surroundings. It enters into the soil
332 and water through diverse pathways like leaking the pipelines, car wash stations, auto workshops
333 and oil purifying factories. There are various reports of biodegradation of diesel by the
334 action of different bacterial species that are found in very harsh conditions, Sorkoh *et al.*,
335 (1993). In addition to the current research (Ijah and Antai, 2003) stated that *Bacillus* species
336 was the most prominent isolate of all diesel fuels utilizing bacteria. As Zhuang *et al.*, (2002)
337 demonstrated that gram negative bacteria allow them to maintain in the extreme variable
338 diesel contaminated environment. In the present exploration three bacterial species namely
339 *Bacillus subtilis*, *Bacillus cereus* and *Pseudomonas aeruginosa* were isolated and they showed the
340 potential of degrading commercial diesel under laboratory conditions.

341 The diesel degrading capability of microbes can be evaluated by using special methods and
342 conventional instruments. Several scientists used Gas chromatograph, GCMS and IR
343 spectroscopy for degradation evaluation. The isolation and augmentation of such microbes on
344 selective media along with commercial diesel has also been considered. In the present work
345 FTIR spectroscopy of Shimadzu Corporation has been used to qualitatively evaluate the
346 determination of commercial diesel fuel presented in two different concentration low and high

347 1000 ppm and 5000 ppm respectively for 15 days. The FTIR analysis showed that the
348 incorporation of bacterial species that was isolated indicate the degradation of diesel fuel as
349 compared with the samples that were not supplemented with bacteria. The results of IR analyses
350 of TPH shows that the additions of all bacteria strains enhance degradation of diesel fuel
351 compared to the sample that was not supplemented with any bacteria strain Yousefi Kebria *et al.*,
352 (2009).

353 The results of the FTIR spectrum were plotted on a graph paper. The spectrum taken for a control
354 sample having diesel is given in Figure 4.4. It obviously indicates the qualitative analysis of
355 sample. It showed IR spectroscopic peak at 2852.72 cm^{-1} , 2922.16 cm^{-1} , 2853.02 cm^{-1} corresponds to
356 the stretching vibration of hydrogen and carbon bonds which was recorded on a graph paper
357 (Figure 4.4). The intensity of the above peaks in the spectrum of control was compared with the
358 intensity of the peaks in the spectrum of the test samples. This response was used as control for
359 comparison with results of test samples offered diesel at a concentration of 1000 ppm. An almost
360 complete removal of diesel components was seen from the reduction in hydrocarbon peaks
361 observed using Solid Phase Micro extraction Gas Chromatography analysis after 5 days of
362 incubation Shukor *et al.*, (2009). The 1000 ppm concentration control intensity of peaks at
363 2852.72 cm^{-1} , 2922.16 cm^{-1} , 2853.02 cm^{-1} was 10.1, 7.11, 4.22 upon calculations the overall intensity
364 was 21.42 which was high as compared with the intensity of a spectrum of the samples. It has
365 been found that only 4 samples showed degradation of commercial diesel as the IR spectroscopic
366 peaks at 2852.72 cm^{-1} , 2922.16 cm^{-1} , 2853.02 cm^{-1} have low intensity as compared with control. In
367 some recent studies done in different countries confirmed the presence of the same bacterial
368 species in different contaminated soil samples as Yousefi Kebria *et al.*, (2009) examined the
369 biodegradation of diesel by *B.cereus* and *B.thuringiensis* strains in 500 and 10000 ppm. It was

370 found that bacterial species degraded the diesel with increasing the concentration of diesel. In the
371 present study *B.cereus* and *B.subtilis* was also isolated and showed the capacity of diesel
372 degradation in the concentration of 1000 ppm and 5000 ppm.

373 Several investigators have reported that inoculation of microbes that acquire the degradative
374 capacity of natural pollutants in cultures may not succeed to function when introduced into the
375 natural environment. With special fertilizers the nutrient sources had a significant collision on
376 diesel degradation of the same isolate. It might be sufficient to harbor microorganisms from a
377 contaminated site, because the microbes have adapted to a contaminated environment and
378 utilizes the contaminant as a sole of carbon and energy (Sohal and Srivastava, 1994; Watanabe,
379 2002; Ghazali *et al.*, 2004; Das and Mukherjee, 2006).The results strongly indicate that the
380 environmental conditions including physical and chemical conditions of the contaminated sites
381 play a crucial role in the degradation even though additional diesel-degrader has been introduced
382 into the contaminated site.

383 The influence of environmental factors rather than genetic capability of a microorganism have
384 been reported to limit the degradation of pollutants.

385 The ability of isolated species *Bacillus subtilis*, *Bacillus cereus* and *Pseudomonas aeruginosa* as
386 bio-degraders of diesel could be very useful for bioremediation of such petroleum hydrocarbon
387 in areas of Mansehra and other localities in Pakistan.

388 **CONCLUSION**

389 We conclude that diesel-degrading bacteria or other oil degrading bacteria are richly found in
390 the oil contaminated soils in Mansehra and this can be demoralized for large oil-spill clean-up
391 campaigns. This study also provides information about the physico-chemical and biological

392 requirements for optimum degradation of commercial diesel by these bacteria in the laboratory
393 condition. The microbes isolated from oil contaminated soils in Mansehra were proficient of
394 degrading diesel with a concentration of 1000 ppm and 5000 ppm. All three species were
395 competent to degrade diesel with variations. Among the isolated organisms the *Pseudomonas*
396 *aeruginosa* were found to degrade diesel up to 1000 ppm and the other isolates namely
397 *Bacillus subtilis*, *Bacillus cereus* degraded diesel best at 1000 ppm rather than 5000 ppm.
398 The study purely describes that both gram positive and gram negative bacteria have the
399 capability to degrade diesel fuel. Where ever we use the bacterial consortium in the natural
400 contaminated environment it should be give better results instead of other chemical process.

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